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A search for myosin in elongating hyphae of *Neurospora crassa*

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Abstract

van Tuinen D., Ortega Perez R. and G. Turian. 1986. A search for myosin in elongating hyphae of *Neurospora crassa*. Bot. Helv. 96: 299–302

Myosin light-chain from smooth muscle has been phosphorylated by a Ca^{2+} , calmodulin-dependent protein kinase from *N. crassa*. Evidence for a myosin-like protein based on the Ca^{2+} -ATPase activity in the absence of Mg^{2+} has been obtained in growing mycelia.

Introduction

Myosin-like proteins are still poorly known in plants (Jackson 1982). In Fungi, they have only been isolated and characterized, but in much smaller amounts than the other major contractile protein actin, from slime molds (Adelman and Taylor 1969) and, recently, from budding yeast cells (Watts et al. 1985). These proteins have several properties in common with muscle myosin as they not only bind to actin into actomyosin but present Ca^{2+} ions activated ATPase activity (Korn 1978).

We have recently proposed a model of polar exocytosis of vesicles in outgrowing germ tubes and elongating hyphae of *Neurospora crassa* implicating the actomyosin system. Actin microfilaments were cytochemically evidenced, but the presence of myosin was only indirectly inferred from the myosin-kinase activity of our Ca^{2+} -calmodulin extracts of *N. crassa* hyphae (van Tuinen 1985). We now present such results corroborated by the first evidence for a myosin-like protein in *N. crassa* founded on the activity of a Ca^{2+} -ATPase in extracts of its elongating hyphae.

Materials and methods

Neurospora crassa strain STA4 (FGSC) has been grown in Vogel's medium (1956) supplemented with 2% sucrose as previously described (Ortega Perez et al. 1981) and the harvested mycelia were frozen in liquid nitrogen and stored at -80°C until use.

Frozen mycelia were disrupted in a blender (Moulinex, type 320) for 2 min. The dry powder was resuspended in 2 vol. extraction buffer: 20 mM imidazole-HCl pH 7.5, 370 mM sucrose,

10 mM β -mercaptoethanol, 10 mM benzamidine, 0.25 μ g/ml pepstatin, and retreated as described above.

The homogenate was filtered through a single layer of nylon cloth and ultracentrifuged at 160,000 g for 90 min in a TFT. 7038 rotor (Prepspin, Kontron). The pellet was resuspended in the extraction buffer supplemented with 1% Triton X 100 and recentrifuged for 60 min as described above. The supernatant obtained was dialysed against the extraction buffer, in which the protease inhibitors were omitted. This dialysate was used for the ATPases assays. All the operations were performed at 4 °C.

The Ca^{2+} , calmodulin-dependent protein kinase has been partially purified by affinity chromatography on a calmodulin-Sepharose column (van Tuinen et al. 1984). The myosin light chain (MLC) of chicken geaser was a gift of Dr. F. Martin, University of Montpellier. The phosphorylation assays were performed at 37 °C in Tris-HCl buffer pH 7.5, containing 5 mM MgCl_2 , 10 mM NaF, 1 mM β -mercaptoethanol and either 3 mM EGTA alone or 3 mM EGTA and 3.5 mM CaCl_2 . Assays of ATPase activities were performed at 37 °C in 250 μ l aliquots containing 50 mM imidazole pH 7.5, 1 mM ATP and when stated 5 mM CaCl_2 or 5 mM MgCl_2 or 2 mM EGTA, 2 mM EDTA and 500 mM KCl. The enzymatic reactions were initiated by adding ATP and stopped by boiling. The P_i was measured as according to Chen et al. (1956).

The phosphorylative reactions were initiated by adding 1 μ Ci [γ - ^{32}P]ATP (3000 Ci/mmol, Amersham Radiochemical Center) and stopped by boiling and then analyzed by electrophoresis on SDS-polyacrylamide gels (Laemmli 1970) followed by autoradiography. The relative incorporations were determined by densitometric analysis of the autoradiograms.

Protein concentrations were determined as described by Spector (1978), using bovin serum albumin as standard.

Results

a) Phosphorylation of myosin light chain by Ca^{2+} , calmodulin-dependent kinase

Following our isolation of a protein kinase from hyphae of *N. crassa*, we have searched for its protein targets (van Tuinen et al. 1984). After detection of an endogenous peptide determined to have molecular weight of 47 kDa, we have tested some other exogenous proteins for their ability to be phosphorylated in a calcium-calmodulin depending manner. Casein, phosphorylase b, histone, microtubule-associated

Table 1. Phosphorylation of MLC by Ca^{2+} , calmodulin-dependent protein kinase from *Neurospora crassa*

	Ca^{2+} CaM	Ca^{2+}	EGTA CaM	Ca^{2+} CaM	Ca^{2+} CaM
Protein kinase	—	+	+	+	+
MLC	+	+	+	+	—
Relative P_i incorporation in the peptide:					
47 kDa	—	32	20	100	176
MLC	0	35	26	45	—

The phosphorylation assays were performed at 37 °C in 45 μ l aliquots containing 20 mM Tris-HCl pH 7.5, 5 mM MgCl_2 , 3 mM EGTA (minus calcium) and 3.5 mM CaCl_2 (plus calcium), 0.3 M KCl, 1 μ Ci ATP and were stated 1 μ g calmodulin (CaM). The EGTA eluate of the calmodulin-Sepharose column contained the protein kinase and the 47 kDa peptide.

Table 2. ATPase activity in the Triton X 100 soluble, supernatant preparation of *Neurospora crassa*

	No addition	K ⁺ /EDTA	CaCl ₂	MgCl ₂
Activity nmol/min · mg	≤ 5	36	399	210

The ATPase assays were performed at 37°C in 250 µl aliquots containing 50 mM imidazole/HCl pH 7.5, 1 mM ATP and one of the following (1) 0.5 M KCl, 2 mM EDTA, 2 mM EGTA; (2) 5 mM CaCl₂, 2 mM EDTA; (3) 5 mM MgCl₂, 2 mM EGTA.

protein, glycogen synthase were not phosphorylated by the Ca²⁺, calmodulin-dependent protein kinase. In contrast, the light chain myosin (MLC) from chicken geaser could efficiently be phosphorylated by a protein kinase eluted from calmodulin-Sepharose affinity column (Table 1). The phosphate incorporation in the MLC was stimulated by a factor of 1.3 by calcium alone and 1.7 by the presence of both calcium and calmodulin with the activity increased in the presence of calmodulin but in the absence of calcium (EGTA).

b) Ca²⁺-ATPase activity

The Ca²⁺ requirement is known to be characteristic for one of the myosin ATPase activities. Therefore we analyzed ion requirements for the ATPase activity present in the ultracentrifuged supernatant, obtained as described above, of *N. crassa* supernatant.

The ATPase activity in the presence of CaCl₂ is about twice as high as the activity observed in the presence of MgCl₂ alone (Table 2). On the other hand, the activity in the presence of K⁺/EDTA is relatively low. A low activity in the presence of K⁺/EDTA has also been observed in *Saccharomyces cerevisiae* (Watts et al. 1985), but in opposition to *N. crassa*, the myosin ATPase activity in yeast was higher in the presence of MgCl₂ than in the presence of CaCl₂.

Discussion

Our ATPasic myosin-like activity will need further characterization to be related to a true fungal myosin, presumably microfilamentous and therefore able to interact with actin microfilaments productive of the shearing force required for the polarized vesicular traffic in the hyphal tips (Turian et al. 1985).

Such interaction could be related to the Ca²⁺-calmodulin dependent myosin-kinase activity detected in elongating hyphae of *N. crassa*, in contrast to higher plants, where MLC from smooth muscle was not phosphorylated by protein kinase (Polya et al. 1983).

In order to exert a mechanical force, the contractile assembly – actin and myosin filaments – must somehow be anchored to other cellular components such as cell membranes (Alberts et al. 1983). The association of our myosin-like protein with the particulate fraction could therefore be explained by the binding of myosin to membranes.

Myosin is the only actin-associated protein that can generate mechanical force derived from ATP hydrolysis (Fulton 1984). Anticipating on its detection in outgrowing germ tubes and elongating hyphal tips of *N. crassa*, we have proposed a model for such an ATP-derived mechanical force for the directional driving of vesicles or exocytosis to the acidified polarly expanding tips (Turian et al. 1985). The first evidence of myosin-like protein in *N. crassa* growing hyphae reinforces our proposal.

Résumé

La chaîne légère de la myosine de muscle lisse a été phosphorylée par une protéine Ca^{2+} , calmoduline-dépendante de *N. crassa*. Dans le mycélium en croissance, la présence d'une protéine de type myosine a été suggérée par l'activité Ca^{2+} -ATPasique obtenue en l'absence d'ions Mg.

Literature

- Adelman M. R. and Taylor E. W. 1969. Further purification and characterization of slime mold myosin and actin. *Biochemistry* 8: 4976–4988.
- Alberts B., Bray D., Lewis J., Raff M., Roberts K. and Watson J. D. 1983. *Molecular biology of the cell*. Garland Publ. Inc. New York, London. 1146 pp.
- Chen P. S., Toribara Jr. T. Y. and Warner H. 1956. Microdetermination of phosphorus. *Anal. Chem.* 28: 1756–1758.
- Fulton A. B. 1984. *The cytoskeleton. Cellular architecture and choreography*. Chapman and Hall, New York, London. 172 pp.
- Jackson W. T. 1982. Actomyosin. In: *The cytoskeleton in plant growth and development*. C. W. Lloyd ed. Acad. Press, London, New York. 457 pp.
- Korn E. D. 1978. Biochemistry in actomyosin-dependent cell motility (a review). *Proc. Natl. Acad. Sci. USA* 75: 588–599.
- Laemmli U. K. 1970. Cleavage of structural protein during the assembly of the head of bacteriophage T4. *Nature* 227: 680–685.
- Ortega Perez R., van Tuinen D., Marmé, D., Cox J. A. and Turian G. 1981. Purification and identification of calmodulin from *Neurospora crassa*. *FEBS Lett.* 133: 205–208.
- Polya G. M., Davies J. R. and Micucci V. 1983. Properties of calmodulin activated Ca^{2+} -dependent protein kinase from wheat germ. *Biochem. Biophys. Acta* 761: 1–12.
- Spector T. 1978. Refinement of the Coomassie blue method of protein quantitation. *Anal. Biochem.* 86: 142–146.
- Turian G., Ton-That T. C. and Ortega Perez R. 1985. Acid tip linear growth in fungi: requirements for $\text{H}^+/\text{Ca}^{2+}$ inverse gradients and cytoskeleton integrity. *Bot. Helv.* 95: 311–322.
- van Tuinen D. 1985. Recherche sur les protéines kinases calcium, calmoduline-dépendantes chez *Neurospora crassa*. Thèse N° 2155, Université de Genève.
- van Tuinen D., Ortega Perez R., Marmé D. and Turian G. 1984. Calcium, calmodulin-dependent protein phosphorylation in *Neurospora crassa*. *FEBS Lett.* 176: 317–320.
- Vogel H. J. 1956. A convenient growth medium for *Neurospora*. *Microbiol. Genet. Bull.* 13: 42–43.
- Watts F. Z., Miller D. M. and Orr E. 1985. Identification of myosin heavy chain in *Saccharomyces cerevisiae*. *Nature* 316: 83–85.